

Amendments to the Specification

Please replace the paragraph at page 8, lines 16-18 with the following amended paragraph:

Figure 10 illustrates the typical organisation of an immunodominant peptide domain for HLA class I-associating peptides whereby several peptide copies arranged in a contiguous fashion are fused to a translocating domain (D) and optionally a C-terminal KDEL (SEQ ID NO: 9)-like sequence.

Please replace the paragraph at page 10, lines 24-28 with the following amended paragraph:

All other DNA manipulation steps were performed according to Sambrook et al. (Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, second edition, 1989). For the insertion of peptide sequences, synthetic oligonucleotides were synthesized using an Applied Biosystems 430A synthesiser and purified by reverse phase HPLC. Sequences were as follows (~~Seq ID No.s 1 to 6 respectively~~);

Please replace the previously amended paragraph at page 11, lines 5-13 with the following amended paragraph:

The oligonucleotides P53-U and -L (encoding the p53 CTL reactive peptide KYICNSSCM (SEQ ID NO: 7) ~~SEQ ID NO: 7~~ (Noguchi *et al.*, 1994 Proc. Natl. Acad. Sci. USA 91, 3171-3175) and FLU-U and -L (encoding the influenza A matrix protein peptide GILGFVFTL (SEQ ID NO: 8) ~~SEQ ID NO: 8~~ reactive with influenza specific CTLs (Gammon *et al.*, 1992 J Immunol. 148, 7-12) were 5'-labelled with ^{32}P using polynucleotide kinase and ^{32}P ATP and annealed together, self-ligated at 37°C for 4 hours

using T4 DNA ligase (Life Technologies, Paisley UK) and analysed on a preparative polyacrylamide sequencing gel. The bands at 180 base pairs representing 5 self-ligated copies of P53-U/L or FLU-U/L were purified, ligated to phosphorylated *NotI* linkers (#1127, New England Biolabs, Hitchin, UK) and digested with *NotI* (Pharmacia).

Please replace the previously amended paragraph at page 11, line 23 to page 12, line 9 with the following amended paragraph:

For preparation of cytotoxic T lymphocytes (CTLs) specific for the p53-derived peptide as above, the *in vivo* mouse peptide immunisation and sensitisation procedure of Noguchi et al. (loc. cit.) was followed to produce long-term CTL lines. For testing of antibodies for ability to induce CTL activity, target mouse Sp2/0 cells (ATCC CRL-1581) were used and maintained in DMEM and 10% foetal bovine serum. For CTL assays, cells at 5×10^5 cells/ml were labeled overnight with 20 μ Ci (7.4 MBq) ^{51}Cr chromate. Cells were then pelleted, washed in medium and resuspended at 5×10^5 cells/ml in medium plus dilutions of the antibody fragments or 10 μ g/ml peptide KYICNSSCM (SEQ ID NO: 7) SEQ ID NO: 7 ("p53 peptide only") for 4 hours at 37°C. Cells were then repelleted, washed twice in PBS (phosphate-buffered saline) and plated at 5×10^3 cells in 100 μ l in RPMI1640 medium plus 10% foetal bovine serum in 24-well plates. 100 μ l of CTLs were then added to give effector:target ratios of 20:1, 10:1 and 5:1 and incubated for 4 hours at 37°C. After incubation, 100 μ l of culture supernatant was carefully removed from each well into an eppendorf tube, centrifuged and triplicate 20 μ l aliquots of supernatant were counted in a scintillation counter. Percent specific release was calculated as [(release by effector cells-spontaneous release)/(maximum release-spontaneous release)] x100. Results were as follows;

Please replace the previously amended paragraph at page 13, lines 10-14 with the following amended paragraph:

Human cytotoxic T lymphocyte (CTLs) specific for the flu peptide GILGFVFTL (SEQ ID NO: 8) ~~SEQ ID NO. 8~~ were obtained from a normal HLA-A2 donor and were maintained as described by Bednarek *et al.*, (1991 *J. Immunological Methods* 139, 41-47). Testing of antibodies for ability to induce CTL activity against target MCF7 cells was as for example 1 with effector:target ratios of 40:1, 20:1 and 10:1. Results were as follows;

Amendments to the Sequence Listing

Please replace all previously filed sequence listings with the pages marked
“Sequence Listing” submitted herewith.